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Scientifically proven to be the most effective instrument pre-clean gel

AZO™ GEL

Instrument Pre-clean Gel

Significant risks have been associated with inadequate or improper cleaning of reusable medical devices[1]. Any soil left on a device following cleaning can put a patient at risk for healthcare associated infections.

The cleaning of medical instruments is a multi-step process. It involves medical personnel handling the instruments post procedure to staff cleaning and preparing it for sterilisation at the decontamination site.

It is essential that practices and processes applied to thorough decontamination of all surgical instruments are one of the highest quality and reflect modern day standards.[2]

The emergence of infectious micro-organisms such as prions, has resulted in a change in practice to prevent and manage the potential spread of diseases such as CJD. Research and evidence support changes such as – moistening of instruments post procedure, streaming of instruments or strategies to prevent migration of instruments.[3]

The importance of keeping instruments moist prior to decontamination has been acknowledged by a number of decontamination guidelines e.g. NICE Interventional Procedure Guidance 196, CFPP 01-01 - Management and Decontamination of Surgical Instruments (Medical Devices) Used in Acute Care, SHEA Guideline for Disinfection and Sterilization of Prion-Contaminated Medical Instruments or AORN Recommended practices for cleaning and caring for surgical instruments and powered equipment.

> “Do not allow instruments to become dry after use. Dried films of tissue are more resistant to prion inactivation by steam sterilisation than are tissues that have been kept moist. Keep instruments moist (…) after use and during storage or transport prior to decontamination in central processing departments.[4]”
>
> SHEA

> “Keeping the environment around soiled instruments at or near saturation humidity (moist) prevents full attachment of hydrophobic proteins such that they are more efficiently removed by cleaning.[5]”
>
> CFPP 01-01

- The prevention of infection is one of the fundamental principles of patient care, underpinning many of the standards, policies and procedures within the perioperative area.[6]
- Keeping medical instruments moist after use and before decontamination is a part of the prevention and control of the risk of transmission of infection through medical instruments.[7]
- Healthcare professionals bear the responsibility for patient safety and for the maintenance of surgical instruments and have a significant role in the decontamination process.

GEL

Product Overview

AZO™ GEL – an advanced pre-clean gel for surgical and dental instruments by, Synergy Health, features an innovative formulation preventing surgical soil & debris from drying onto instruments.

Key Features & Benefits

✓ Scientifically advanced, patent pending formulation	AZO™ GEL facilitates cleaning of reusable surgical and dental instruments, increasing productivity of decontamination sites[8]
✓ Optimal adhesion to surface ✓ Most stable coating of the instruments	AZO™ GEL reduces spills & splashes, protecting personnel handling contaminated instruments[9]
✓ Neutral pH ✓ Contains corrosion inhibitor ✓ Broad material compatibility	AZO™ GEL protects your instruments[10]
✓ No dyes ✓ No surfactants ✓ No fragrance	AZO™ GEL is compatible with washer disinfector systems

Hospital Product Evaluation Trial

Trial Site The London Clinic, London

Trial Duration 4 weeks, March 2013

User acceptability feedback from operating theatre staff indicates that:

- **100%** respondents find AZO™ GEL easy to use
- **86%** staff acceptability of the AZO™ GEL
- **71%** respondents have indicated that the product meets their expectations

Sterilisation site reported operational benefits in reprocessing of surgical instruments

21% increase in instruments processed per hour

36% reduction in re-washes

"AZO™ GEL is easy to use and it does not irritate me as the product we currently use."

"The gel is working very well; we need to apply it to all trays."
"No extra work is required to remove dirt or debris."

Objective

Determine the effectiveness of different instrument pre-cleaners in adhering onto material surface.

Methodology

The test plates were made of AISI 316L stainless steel measured 100 x 160 x 3 mm thick. The plates were mechanically polished down to a dull polish finish prior to cleansing with soapy water and degreasing with ethanol in ultrasonic bath for 15 minutes. The plates were then dried prior to use.

Competitor products A to D represent brands currently used in the market and were sourced from independent distributors.

All tests were performed at 18°C. The plates were placed in horizontal position and sprayed with approximately 7 ml of test solution to achieve uniform coating. The plates were then raised into vertical position for 10 minutes. At every 2 minute interval, photographs were taken to determine the position of the red tabs. The adhesion test was repeated three times.

Observations

After 10 minutes in the vertical position, 100% of AZO™ GEL remains on the stainless steel surface. Only 17% of Product A remains on the surface. Products C – D have completely drained off by the 2 minute interval.

Conclusion

AZO™ GEL provides the most stable coating on stainless surfaces in comparison to four other products currently used in the market.

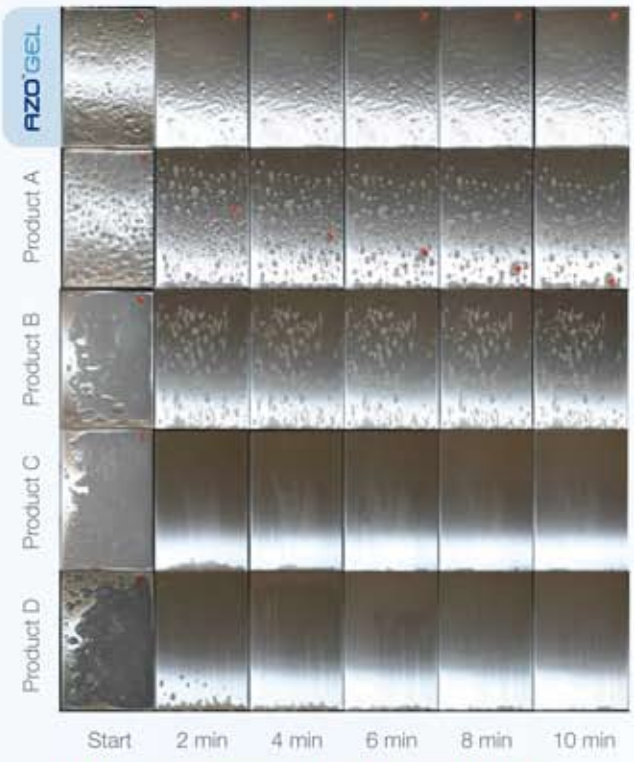


Fig 1 Surface adhesion test - ENSPEC UK Ltd

Fig 2 After 10 minutes in vertical position, 100% of AZO™ GEL remains on the surface of the test plate

Fig 3 The plates of products C & D completely drained off after 2 minutes

Objective

Determine the effectiveness of different instrument pre-cleaners in soil removal.

Methodology

The test plates were cleaned and processed using the same procedure adopted for adhesion tests.

Competitor products A to D represent brands currently used in the market and were sourced from independent distributors.

All tests were performed at 18°C. The test plates were prepared by placing three 0.5 ml drops of Browne test soil spaced evenly, near the top of plates and left to dry in the horizontal position for 30 minutes. The test plates were then sprayed with approximately 7 ml of test solution to achieve uniform coating, and raised to the vertical position for 30 minutes. At the end the test plates were rinsed for 10 seconds using a low pressure water jet across the soil spots and plate surfaces to determine the ease of removing dried soil deposit on the stainless surfaces.

Observations

With light rinsing for 10 seconds, there is a bit of one Browne drop remaining on the plate treated with AZO™ GEL whilst all other products A – D still have dried matter (three drops of Browne soil) attached to them.

Conclusion

The observations suggest that AZO™ GEL is most effective in aiding the removal of dried soil from stainless steel surfaces as compared to four other products currently used in the market.

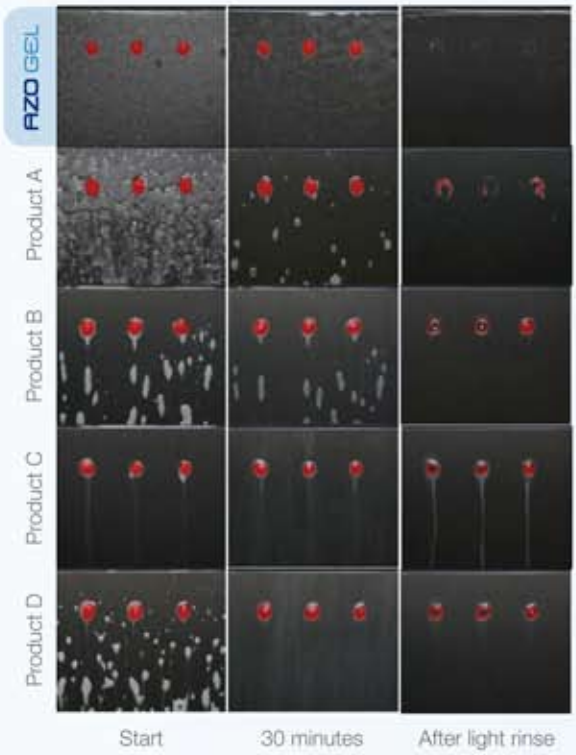


Fig 4 Soil lifting test - ENSPEC UK Ltd

Fig 5 Surgical instrument with dried on blood

Fig 6 30 seconds after AZO™ GEL application blood is lifted off the surface of the instrument

1

Place soiled instruments in tray.

2

Ensure all instruments are open.

3

Place tray in plastic bag.

4

Spray directly onto soiled instruments.

5

Ensure all instruments are well coated.

6

Close the bag.

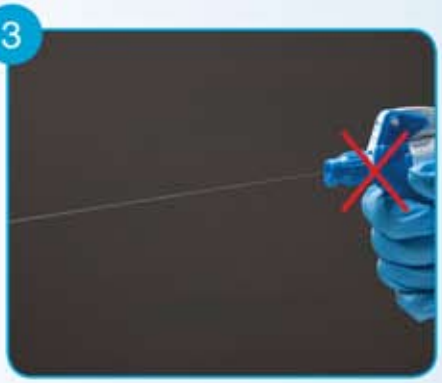
1

Turn the nozzle anti-clockwise.

2

Depress trigger fully for a fine mist. This will ensure optimal coating of the instruments.

3



Do not turn trigger spray nozzle too far. This will produce a stream of gel.

4

Incomplete actuation will result in stream of gel or spray releasing droplets.

5

If paper bags are in use with dental instrument trays, coat dental instruments prior to placing dental tray in paper bag.

6

Pay special attention to ribbed, knurled, etched and heavily soiled surfaces. Coat them well with AZO™ GEL.

Materials Compatibility

AZO™ GEL

AZO™ GEL has been tested by experts in corrosion against a wide range of materials used in surgical and dental instruments. It is safe for use on and will not corrode the following metals:

- ✓ Polycarbonate
- ✓ Polypropylene (PP)
- ✓ Polyethylene (PET)
- ✓ Polymethyl methacrylate (PPMA)
- ✓ Polytetrafluoroethylene (PTFE)
- ✓ Polyether ether ketone (PEEK)
- ✓ Martensitic stainless steel
- ✓ Austenitic stainless steel
- ✓ Aluminium alloy
- ✓ Chrome plated carbon steel
- ✓ Acetal copolymer
- ✓ Polyvinyl chloride (PVC)
- ✓ Brass
- ✓ Gold
- ✓ Nickel
- ✓ Titanium
- ✓ Nylon
- ✓ Nitrile rubber

Ordering Information

Code	Product name	Product description	Size	Case qty
AZG750S	AZO™ GEL	Instrument pre-clean gel	750 ml	6

For more information or to place your order, contact our customer care team on

0800 226 3673

References: 1. Stoessel K, Moore K, Shoemake S. Cleaning Reusable Medical Devices: A Critical First Step. Managing Infection Control May 2007; 74-80 **2.** The Association for Perioperative Practice (AfPP); http://www.afpp.org.uk/news/resources/decontamination **3.** The Association for Perioperative Practice (AfPP); http://www.afpp.org.uk/news/resources/decontamination **4.** William A. Rutala, David J. Weber. Guideline for Disinfection and Sterilization of Prion-Contaminated Medical Instruments. Infection control and hospital epidemiology February 2010, vol. 31; no. 2; 111 **5.** Choice Framework for local Policy and Procedures 01-01 – Management and decontamination of surgical instruments (medical devices) used in acute care. Part A: the formulation of local policy and choices manual version:9949: 1.0:England, August 2012; 57-58 **6.** The Association for Perioperative Practice (AfPP); http://www.afpp.org.uk/news/resources/decontamination **7.** Choice Framework for local Policy and Procedures 01-01 – Management and decontamination of surgical instruments (medical devices) used in acute care. Part A: the formulation of local policy and choices manual version:9949: 1.0:England, August 2012; 11-12 **8.** Internal data on file **9.** Internal data on file **10.** Internal data on file **11.** ENSPEC Technology Technical Report Case No EN2505 Rev2; AZO GEL Materials Compatibility Study 3 May 2013

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